

# TEMPERATURE PROFILES OF SOLUBLE AND BOUND PECTIN METHYLESTERASE FROM APPLE PEEL

Ahmet Yemenicioğlu\*  
Bekir Cemeroglu\*\*

## ABSTRACT

Soluble (SPME) and bound pectin methylesterase (BPME) from Golden Delicious apple peel were separated and partially purified by 0-95%  $(\text{NH}_4)_2\text{SO}_4$  precipitation, dialysis and centrifugation. The specific activities of SPME and BPME were 4.2 and 3.5 folds higher than that of the crude extract. The temperatures for the 50% loss of SPME and BPME in 5 min were 70° for SPME and 67.5°C for BPME, respectively. Both fraction lost almost 90% of their activities in 5 min at 80°C

**Key words:** Apple, pectin methylesterase, purification, temperature profile

## ELMA KABUĞUNDA BULUNAN ÇÖZÜNÜR VE BAĞLI PEKTİN METİLESTERAZ ENZİMLERİNİN SICAKLIK DEREJESİ PROFİLLERİNİN BELİRLENMESİ

### ÖZET

Golden Delicious elmalarında bulunan çözü-  
nür (SPME) ve bağlı pektin metilesteraz

(BPME) enzimleri birbirinden ayrılmış ve %0-90  $(\text{NH}_4)_2\text{SO}_4$  ile çöktürülerek, dializ ve santrifüj uygulanarak kısmi saflaştırma sağlanmıştır. Bu şekilde ayrılmış olan SPME ve BPME enzimlerinin spesifik aktiviteleri, ham ekstraktın aktivitesine göre sırasıyla 4.2 ve 3.5 katına ulaşmıştır. SPME ve BPME enzimlerinin 5 dakika içerisinde aktivitelerinin %50'sini yitirdikleri sıcaklık dereceleri sırasıyla; 70° ve 67.5°C dir. 80°C de 5 dakikalık bir ısıtma, her iki fraksiyonun aktivitelerinin %90'ını yitirmesine neden olmaktadır.

**Anahtar kelimeler:** Elma, pektin metilesteraz, saflaştırma, sıcaklık derecesi profili

### INTRODUCTION

Hot water immersion (HWI) of fruits is a quarantine treatment for the disinfection of fruit flies and prevention of fungal activity (1, 2). The temperature of the HWI treatment should be chosen very carefully to prevent heat damage (3). The suitable temperatures of HWI treatment for some fruits were reported as 45°C (30-60 min) and 46°C (30-50 min) for cucumbers (4), 42-49°C for papaya fruits (2) and 45°C for some

\* Ankara University, Faculty of Agriculture Department of Food Engineering Dışkapı, Ankara. 06110 Turkey.

\*\* To whom correspondence should be addressed.

apple cultivars (1). HWI was also reported to improve the texture of some apple cultivars, i.e. Golden Delicious and Delicious (1).

The increase in firmness by heat treatment is attributed to the accelerated action of Pectin Methylesterase (PME, E.C.3.1.11) which demethylates pectic compounds and causes the increase of their interaction with divalent ions, i.e.  $\text{Ca}^{++}$  and  $\text{Mg}^{++}$  (5, 6, 7). This interaction creates a rigid network, called the egg-box model and improves the texture (8, 9).

The thermal characteristics of in situ PME activity of apple waste (10) and PME from whole apple fruits (11) were investigated. However, no studies have been found related to the PME from apple peel. In this work the temperature profiles of soluble and cell wall bound PME from Golden delicious apple peel were determined for the better understanding of the thermal characteristics of enzyme PME.

## MATERIAL AND METHODS

### Material

Golden Delicious apples were obtained from the experimental orchards of Ankara University.

### Methods

#### Extraction of enzymes

Acetone powder, obtained according to the method of Yemenicioğlu et al (12), was used as enzyme source in this study. All extraction and inactivation studies were carried out in 0.01M Tris-HCl buffer at pH 7.5 unless otherwise stated.

#### Buffer extraction

3-5g acetone powder was suspended in 200 mL cold buffer (+4°C) containing 1M NaCl. The

extraction was carried out 24h at +4°. The slurry was then filtered through a muslin cloth and centrifuged at 7000g for 1h.

#### $(\text{NH}_4)_2\text{SO}_4$ precipitation and dialysis

For further purification the supernatant was brought to 0-95%  $(\text{NH}_4)_2\text{SO}_4$  saturation and stirred 3.5h at +4°C. The formed precipitate was separated by centrifugation at 7000g for 1.5h, dissolved in minimum amount of buffer and dialysed 48h against the same buffer by two changes (2 x 2L). After dialysis, the formed precipitate was collected by centrifugation and the clear supernatant was named as Soluble PME (SPME).

#### Preparation of bound PME extract

On this purpose the precipitate separated from the centrifugation of dialyzate was washed with 3 mL of distilled water (+4°C) and dissolved in 3 mL of buffer. This extract was named as bound PME extract (BPME).

#### Determination of protein

The protein was determined according to the method of Lowry (13).

#### PME activity

PME activity was determined titrimetrically by the method of Alonso et al (8) with modifications. The reaction mixture consisted of 0.2-0.5 mL of PME extract and 20 mL of 0.5% citrus pectin (Sigma Chem. Co. St Louis, MO) solution in 0.1N NaCl. The pH of the mixture was immediately adjusted to 7.5 by adding 0.1N NaOH and maintained at this value for 5 min by titrating slowly with 0.01N NaOH. The titration was carried out in a cell maintained at 30°C constant temperature and the activity was expressed as

units (U). One unit of PME activity was defined as the  $\mu\text{M}$  of carboxyl groups released by 1 mL of enzyme extract in 5 min.

### Temperature profiles

For the determination of temperature profiles, 0.3 mLs of PME extracts were incubated in thermal inactivation time (TIT) tubes (i.d. 9 mm, wall thickness 1 mm) for 5 min at various temperatures (50-70°C). The percent PME activity remaining after each heat treatment was calculated from the initial activity.

## RESULTS AND DISCUSSION

### Partial purification

The crude extract of acetone powder from golden Delicious apples contained very low PME activity (90110U. mL<sup>-1</sup>). The total activity of SPME was almost two-fold higher than that of BPME (Tab. 1). The respective specific activities of SPME and BPME were 4.2 and 3.5 folds higher compared to that of crude extract.

### Temperature profiles

The temperature profiles of SPME and BPME are given in Fig. 1. A linear temperature profile was obtained for BPME whereas, SPME showed gradual increase in activity from 50° to 60°C and a sharp decrease above 60°C. Comparing to the temperature profile of BPME, the SPME inactivation was found to be more affected by the increase of temperature.

The temperatures for the 50% loss of PME activity in 5 min were 67.5°C for BPME and 70°C for SPME, respectively. Both BPME and SPME lost almost 90% of their activities in 5 min at 80°C.

Comparison of our heat inactivation data revealed that PME from apple peel was more heat stable than PME from Bramley apple [ $t_{1/2} \cong 5$  min at 60°C] (10), Purified PME from mandarin orange [ $t_{1/2} \cong 1$  min at 65°C] (14) and PME from Valencia orange [ $t_{1/2} \cong 0.5$  min at 70°C] (15). Therefore, temperatures applied in quarantine treatment [40-50°C (1)] is not likely to be very effective on the inactivation of PME from G. Delicious apple peel.

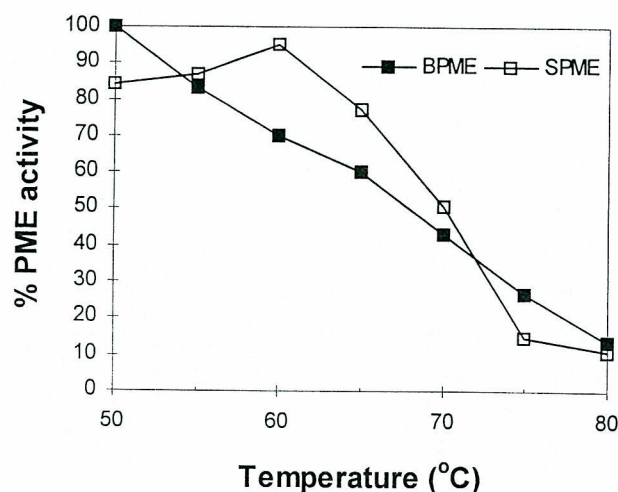
### References

- (1) Kim, D.M., Smith, N.L. and Lee, C.Y. 1993. Apple cultivar variations in response to heat treatment and minimal processing. *J. Food Sci.* 58: 1111-1114, 1124.
- (2) Chan, H.T., Maindonald, J.M., Laidlaw, W.G. and Sellenrich, M. 1996 ACC oxidase in papaya sections after heat treatment. *J. Food Sci.* 61:1182-1185, 1190.
- (3) Underhill, S.J.R. and Critchley, C. 1993. Lychee pericarp browning caused by heat injury. *Hort. Sci.* 28:721-722.
- (4) Chan, H.T. and Linse E. 1989. Conditioning cucumbers to increase heat resistance in the EFE system. *J. Food Sci.* 54: 1375-1376.
- (5) Hudson, J.M. and Buescher, R.W. 1986. Relationship between degree of pectin methylation and tissue firmness of cucumber pickles. *J. Food Sci.* 51: 138-140, 149.
- (6) Stanley, D.W., Bourne, M.C., Stone, A.P. and Wismer, W.V. 1995. Low temperature blanching effects on chemistry, firmness and structure of canned beans and carrots. *J. Food Sci.* 60: 327-333.
- (7) Verlinden, B.E. and De Beardemaeker, J. 1997. Modeling low temperature blanched carrot firmness based on heat induced processes and enzyme activity. *J. Food Sci.* 62: 213-218, 229.
- (8) Alonso, J., Rodriguez, T., and Canet, W. 1995. Effect of calcium pretreatments on the texture of frozen cherries. Role of pectinesterase in the pectic materials. *J. Agric. Food Chem.* 43: 1011-1016.
- (9) Tang, H.L. and Mcfeeters. R.F. 1983. Relationship among cell wall constituents, calcium and texture during cucumber fermentation and storage. *J. Food Sci.* 48: 66-70.
- (10) King, K. 1990. Partial characterization of the in situ activity of pectinesterase in bramley apple. *Int. J. Food Sci. and Tech.* 25: 188-197.

- (11)Castaldo, D., Quagliuolo, L., Servillo, L., Balestrieri, C. and Giovane, A. 1989. Isolation and characterization of Pectin Methylsterase from apple fruit. *J. Food Sci.* 54:653-655,673.
- (12)Yemenicioğlu, A., Özkan, M. and Cemeroğlu, B. 1997. Heat inactivation kinetics of apple polyphenoloxidase and activation of its latent form. *J. Food Sci.* 62:508-510.
- (13)Harris, D.A. 1987. Spectrophotometric assays. In *spectrophotometry and, spectrofluorometry*, D.A. Harris and C.L. Bashford (Eds.), I.R.L. Press, Oxford. p.49.90.
- (14)Rillo, L., Castaldo, D., Giovane, A., Servillo, L., Balestrieri, C. and Quagliuolo, L. 1992 Purification and properties of pectin methylsterase from mandarin orange fruit. *J. Agric. Food Chem.* 40: 591-593.
- (15)Wicker, L., Vassallo, M.R. and Echeverria, E.J. 1988. Solubilization of cell wall bound, thermostable pectinesterase from valencia orange *J. Food Sci.* 53: 1171-1174, 1180.

**Table 1– Partial purification and separation of soluble and bound PME from apple peel**

Fraction	Volume (mL)	T. Activity (Unit. $10^{-4}$ )	T. Protein (mg)	S. Activity (U. $\text{mg}^{-1}$ ). $10^{-4}$	Purity (Fold)	Recovery (%)
<b>Crude extract:</b>	182	1640	23.7	69.2	1.0	100
<b>0-90% <math>(\text{NH}_3)_2\text{SO}_4</math> precipitation and dialysis:</b>						
Soluble PME	5.9	243	0.832	292	4.2	15
Bound PME	3.0	131	0.535	245	3.5	8.0



**Figure 1- Temperature profiles of soluble and bound pectin methylsterase from apple peel.**